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0305831.0

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855589801

4. Title of the invention

BROAD PROTECTION MENINGOCOCCAL VACCINES

5. Name of your agent *(if you have one)*

Carpmaels & Ransford

"Address for service" in the United Kingdom to which all correspondence should be sent *(including the postcode)*43 Bloomsbury Square
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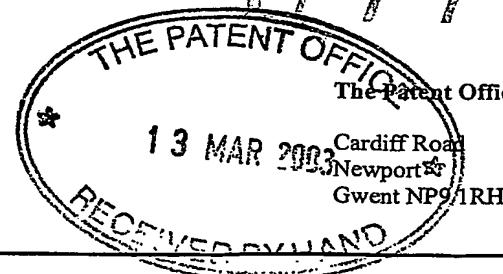
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BROAD PROTECTION MENINGOCOCCAL VACCINES

All documents cited herein are incorporated by reference in their entirety.

TECHNICAL FIELD

This invention is in the fields of immunology and vaccinology. In particular, it relates to antigens 5 from *Neisseria meningitidis* (meningococcus) and their use in immunisation.

BACKGROUND ART

N.meningitidis is a non-motile, Gram-negative human pathogen that colonises the pharynx and causes meningitis (and, occasionally, septicaemia in the absence of meningitis). It causes both endemic and epidemic disease. Following the introduction of the conjugate vaccine against 10 *Haemophilus influenzae*, *N. meningitidis* is the major cause of bacterial meningitis in the USA.

Based on the organism's capsular polysaccharide, various serogroups of *N.meningitidis* have been identified. Serogroup A is the pathogen most often implicated in epidemic disease in sub-Saharan Africa. Serogroups B and C are responsible for the vast majority of cases in the United States and in most developed countries. Serogroups W135 and Y are responsible for the rest of the cases in the 15 USA and developed countries. After serogroup, classification includes serotype, serosubtype and then immunotype, and the standard nomenclature lists serogroup, serotype, serosubtype, and immunotype, each separated by a colon e.g. B:4:P1.15:L3,7,9. Within serogroup B, some lineages cause disease often (hyperinvasive), some lineages cause more severe forms of disease than others (hypervirulent), and others rarely cause disease at all. Seven hypervirulent lineages are recognised, 20 namely subgroups I, III and IV-1, ET-5 complex, ET-37 complex, A4 cluster and lineage 3. These have been defined by multilocus enzyme electrophoresis (MLEE), but multilocus sequence typing (MLST) has also been used to classify meningococci [1].

A polysaccharide vaccine against serogroups A, C, W135 & Y has been known for many years [2, 3] but a vaccine against serogroup B has proved elusive. Vaccines based on outer-membrane vesicles 25 have been tested [e.g. ref. 4], but the protection afforded by these vaccines is typically restricted to the strain used to make the vaccine. There remains a need, therefore, for a broadly-effective serogroup B vaccine.

Genome sequences for meningococcal serogroups A [5] and B [6, 7] have been reported, and the 30 serogroup B sequence has been studied to identify vaccine antigens [e.g. refs. 8 to 13]. Candidate antigens have been manipulated to improve heterologous expression [refs. 14 to 16].

It is an object of the invention to provide further and improved compositions for providing immunity against meningococcal disease and/or infection, and in particular for providing broad immunity against serogroup B meningococcus.

DISCLOSURE OF THE INVENTION

The invention provides a composition which, after administration to a subject, is able to induce an antibody response in that subject, wherein the antibody response is bactericidal against two or more (e.g. 2 or 3) of hypervirulent lineages A4, ET-5 and lineage 3 of *N.meningitidis* serogroup B.

5 Vaccines against pathogens such as hepatitis B virus, diphtheria and tetanus typically contain a single protein antigen (e.g. the HBV surface antigen, or a tetanus toxoid). In contrast, acellular whooping cough vaccines typically contain at least three *B.pertussis* proteins, and the Prevenar pneumococcal vaccine contains seven separate conjugated saccharide antigens. Other vaccines such as cellular pertussis vaccines, the measles vaccine, the inactivated polio vaccine (IPV) and meningococcal
10 OMV vaccines are by their very nature complex mixtures of a large number of antigens.

15 Whether protection against can be elicited by a single antigen, a small number of defined antigens, or a complex mixture of undefined antigens, therefore depends on the pathogen in question. Rather than consisting of a single antigen, it is preferred that the composition of the invention comprises a mixture of 10 or fewer (e.g. 9, 8, 7, 6, 5, 4, 3, 2) purified antigens, and it is particularly preferred that the composition should not include complex or undefined mixtures of antigens e.g. it is preferred not to include outer membrane vesicles in the composition.

20 For serogroup B meningococcus, a mixture of five defined protein antigens has been found to elicit a good protective immune response. The invention thus provides a composition comprising the following five meningococcal protein antigens: (1) a 'NadA' protein; (2) a '741' protein; (3) a '936' protein; (4) a '953' protein; and (5) a '287' protein. These antigens are referred to herein as the 'five basic antigens'.

NadA protein

25 'NadA' (Neisserial adhesin A) from serogroup B of *N.meningitidis* is disclosed as protein '961' in reference 10 (SEQ IDs 2943 & 2944) and as 'NMB1994' in reference 6 (see also GenBank accession numbers: 11352904 & 7227256). A detailed description of the protein can be found in reference 17. There is no corresponding protein in serogroup A [5, 17].

When used according to the present invention, NadA may take various forms. Preferred forms of NadA are truncation or deletion variants, such as those disclosed in references 14 to 16. In particular, NadA without its C-terminal membrane anchor is preferred (e.g. deletion of residues 351-405 for strain 2996 [SEQ ID 1]), which is sometimes distinguished herein by the use of a 'C' superscript e.g. NadA^(C). Expression of SEQ ID 1 in *E.coli* results in secretion of the NadA protein into the culture supernatant with concomitant removal of its 23mer leader peptide (e.g. to leave a 327mer for strain 2996 [SEQ ID 2]). Polypeptides without their leader peptides are sometimes distinguished herein by the use of a 'NL' superscript e.g. NadA^(NL) or NadA^{(C)(NL)}.

Preferred NadA sequences have 50% or more identity (e.g. 60%, 70%, 80%, 90%, 95%, 99% or more) to SEQ ID 2. This includes NadA variants (e.g. allelic variants, homologs, orthologs, paralogs, mutants, etc.). Allelic forms of NadA are shown in Figure 9 of reference 18.

Other preferred NadA sequences comprise at least n consecutive amino acids from SEQ ID 1, 5 wherein n is 7 or more (e.g. 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more). Preferred fragments comprise an epitope from NadA. Other preferred fragments lack one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or the N-terminus of SEQ ID 1 (e.g. NadA^(C), NadA^(NL), NadA^{(C)(NL)}). A preferred fragment of SEQ ID 1 is SEQ ID 2.

10 Secreted NadA can conveniently be prepared in highly pure form from culture supernatant by a process comprising the steps of: concentration and diafiltration against a buffer by ultrafiltration; anionic column chromatography; hydrophobic column chromatography; hydroxylapatite ceramic column chromatography; diafiltration against a buffer; and filter sterilisation. Further details of the process are given in the examples.

15 NadA is preferably used in an oligomeric form.

741 protein

'741' protein from serogroup B is disclosed in reference 10 (SEQ IDs 2535 & 2536) and as 'NMB1870' in reference 6 (see also GenBank accession number GI:7227128). The corresponding protein in serogroup A [5] has GenBank accession number 7379322. 741 is a lipoprotein.

20 When used according to the present invention, 741 protein may take various forms. Preferred forms of 741 are truncation or deletion variants, such as those disclosed in references 14 to 16. In particular, the N-terminus of 741 may be deleted up to and including its poly-glycine sequence (i.e. deletion of residues 1 to 72 for strain MC58 [SEQ ID 3]), which is sometimes distinguished herein by the use of a 'ΔG' prefix. This deletion can enhance expression. The deletion also removes 741's lipidation site.

25 Preferred 741 sequences have 50% or more identity (e.g. 60%, 70%, 80%, 90%, 95%, 99% or more) to SEQ ID 3. This includes 741 variants (e.g. allelic variants, homologs, orthologs, paralogs, mutants, etc.). Allelic forms of 741 can be found in SEQ IDs 1 to 22 of reference 16, and in SEQ IDs 1 to 23 of reference 19.

30 Other preferred 741 sequences comprise at least n consecutive amino acids from SEQ ID 3, wherein n is 7 or more (e.g. 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more). Preferred fragments comprise an epitope from 741. Other preferred fragments lack one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or the N-terminus of SEQ ID 3.

Protein 741 is an extremely effective antigen for eliciting anti-meningococcal antibody responses, and it is expressed across all meningococcal serogroups. The protein groups into three variants, and while serum raised against a given variant is bactericidal within the same variant group, it is not active against strains which express one of the other two variants *i.e.* there is intra-variant cross-protection, but not inter-variant cross-protection. For maximum cross-strain efficacy, therefore, it is preferred that a composition should include more than one variant of protein 741. An exemplary sequence from each variant is given in SEQ ID 10, 11 and 12 herein, starting with a N-terminal cysteine residue to which a lipid will be covalently attached in the lipoprotein form of 741.

It is therefore preferred that the composition should include at least two of: (1) a first protein, comprising an amino acid sequence having at least $a\%$ sequence identity to SEQ ID 10 and/or comprising an amino acid sequence consisting of a fragment of at least x contiguous amino acids from SEQ ID 10; (2) a second protein, comprising an amino acid sequence having at least $b\%$ sequence identity to SEQ ID 11 and/or comprising an amino acid sequence consisting of a fragment of at least y contiguous amino acids from SEQ ID 11; and (3) a third protein, comprising an amino acid sequence having at least $c\%$ sequence identity to SEQ ID 12 and/or comprising an amino acid sequence consisting of a fragment of at least z contiguous amino acids from SEQ ID 12.

The value of a is at least 85 *e.g.* 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99, 99.5, or more. The value of b is at least 85 *e.g.* 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99, 99.5, or more. The value of c is at least 85 *e.g.* 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99, 99.5, or more.

20 The values of a , b and c are not intrinsically related to each other.

The value of x is at least 7 *e.g.* 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 35, 40, 45, 50, 60, 70, 80, 90, 100, 120, 140, 160, 180, 200, 225, 250). The value of y is at least 7 *e.g.* 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 35, 40, 45, 50, 60, 70, 80, 90, 100, 120, 140, 160, 180, 200, 225, 250). The value of z is at least 7 *e.g.* 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 35, 40, 45, 50, 60, 70, 80, 90, 100, 120, 140, 160, 180, 200, 225, 250). The values of x , y and z are not intrinsically related to each other.

It is preferred that any given 741 amino acid sequence will not fall into more than one of categories (1), (2) and (3). Any given 741 sequence will thus fall into only one of categories (1), (2) and (3). It is thus preferred that: protein (1) has less than $i\%$ sequence identity to protein (2); protein (1) has less than $j\%$ sequence identity to protein (3); and protein (2) has less than $k\%$ sequence identity to protein (3). The value of i is 60 or more (*e.g.* 61, 62, 63, 64, 65, 66, 67, 68, 69, 70, 71, 72, 73, 74, 75, 76, 77, 78, 79, 80, 81, 82, 83, 84, *etc.*) and is at most a . The value of j is 60 or more (*e.g.* 61, 62, 63, 64, 65, 66, 67, 68, 69, 70, 71, 72, 73, 74, 75, 76, 77, 78, 79, 80, 81, 82, 83, 84, *etc.*) and is at most b . The value of k is 60 or more (*e.g.* 61, 62, 63, 64, 65, 66, 67, 68, 69, 70, 71, 72, 73, 74, 75, 76, 77, 78, 79,

80, 81, 82, 83, 84, *etc.*) and is at most c . The values of i , j and k are not intrinsically related to each other.

936 protein

5 '936' protein from serogroup B is disclosed in reference 10 (SEQ IDs 2883 & 2884) and as 'NMB2091' in reference 6 (see also GenBank accession number GI:7227353). The corresponding gene in serogroup A [5] has GenBank accession number 7379093.

When used according to the present invention, 936 protein may take various forms. Preferred forms of 936 are truncation or deletion variants, such as those disclosed in references 14 to 16. In particular, the N-terminus leader peptide of 936 may be deleted (*i.e.* deletion of residues 1 to 23 for strain MC58 10 [SEQ ID 4]) to give 936^(NL).

Preferred 936 sequences have 50% or more identity (*e.g.* 60%, 70%, 80%, 90%, 95%, 99% or more) to SEQ ID 4. This includes variants (*e.g.* allelic variants, homologs, orthologs, paralogs, mutants *etc.*).

15 Other preferred 936 sequences comprise at least n consecutive amino acids from SEQ ID 4, wherein n is 7 or more (*e.g.* 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more). Preferred fragments comprise an epitope from 936. Other preferred fragments lack one or 20 more amino acids (*e.g.* 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or the N-terminus of SEQ ID 4.

953 protein

20 '953' protein from serogroup B is disclosed in reference 10 (SEQ IDs 2917 & 2918) and as 'NMB1030' in reference 6 (see also GenBank accession number GI:7226269). The corresponding protein in serogroup A [5] has GenBank accession number 7380108.

When used according to the present invention, 953 protein may take various forms. Preferred forms 25 of 953 are truncation or deletion variants, such as those disclosed in references 14 to 16. In particular, the N-terminus leader peptide of 953 may be deleted (*i.e.* deletion of residues 1 to 19 for strain MC58 [SEQ ID 5]) to give 953^(NL).

Preferred 953 sequences have 50% or more identity (*e.g.* 60%, 70%, 80%, 90%, 95%, 99% or more) to SEQ ID 5. This includes 953 variants (*e.g.* allelic variants, homologs, orthologs, paralogs, mutants, *etc.*). Allelic forms of 953 can be seen in Figure 19 of reference 12.

30 Other preferred 953 sequences comprise at least n consecutive amino acids from SEQ ID 5, wherein n is 7 or more (*e.g.* 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more). Preferred fragments comprise an epitope from 953. Other preferred fragments lack one or more amino acids (*e.g.* 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or the N-terminus of SEQ ID 5.

287 protein

'287' protein from serogroup B is disclosed in reference 10 (SEQ IDs 3103 & 3104), as 'NMB2132' in reference 6, and as 'GNA2132' in reference 13 (see also GenBank accession number GI:7227388). The corresponding protein in serogroup A [5] has GenBank accession number 7379057.

- 5 When used according to the present invention, 287 protein may take various forms. Preferred forms of 287 are truncation or deletion variants, such as those disclosed in references 14 to 16. In particular, the N-terminus of 287 may be deleted up to and including its poly-glycine sequence (*i.e.* deletion of residues 1 to 24 for strain MC58 [SEQ ID 6]), which is sometimes distinguished herein by the use of a ' ΔG ' prefix. This deletion can enhance expression.
- 10 Preferred 287 sequences have 50% or more identity (*e.g.* 60%, 70%, 80%, 90%, 95%, 99% or more) to SEQ ID 6. This includes 287 variants (*e.g.* allelic variants, homologs, orthologs, paralogs, mutants, *etc.*). Allelic forms of 287 can be seen in Figures 5 and 15 of reference 12, and in example 13 and figure 21 of reference 10 (SEQ IDs 3179 to 3184).

Other preferred 287 sequences comprise at least n consecutive amino acids from SEQ ID 6, wherein 15 n is 7 or more (*e.g.* 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more). Preferred fragments comprise an epitope from 287. Other preferred fragments lack one or more amino acids (*e.g.* 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or the N-terminus of SEQ ID 6.

Fusion proteins

- 20 The five antigens may be present in the composition as five separate proteins, but it is preferred that at least two of the antigens are expressed as a single polypeptide chain (a 'hybrid' protein [refs. 14 to 16]) *e.g.* such that the five antigens form fewer than five polypeptides. Hybrid proteins offer two principal advantages: first, a protein that may be unstable or poorly expressed on its own can be assisted by adding a suitable hybrid partner that overcomes the problem; second, commercial manufacture is simplified as only one expression and purification need be employed in order to produce two separately-useful proteins.
- 25

A hybrid protein included in a composition of the invention may comprise two or more (*i.e.* 2, 3, 4 or 5) of the five basic antigens. Hybrids consisting of two of the five basic antigens are preferred.

- 30 Within the combination of five basic antigens, an antigen may be present in more than one hybrid protein and/or as a non-hybrid protein. It is preferred, however, that an antigen is present either as a hybrid or as a non-hybrid, but not as both, although it may be useful to include protein 741 both as a hybrid and a non-hybrid (preferably lipoprotein) antigen, particularly where more than one variant of 741 is used.

Two-antigen hybrids for use in the invention comprise: NadA & 741; NadA & 936; NadA & 953; NadA & 287; 741 & 936; 741 & 953; 741 & 287; 936 & 953; 936 & 287; 953 & 287. Preferred two-antigen hybrids comprise: 741 & 936; 953 & 287.

Hybrid proteins can be represented by the formula $\text{NH}_2\text{-A-[-X-L]}_n\text{-B-COOH}$, wherein: X is an 5 amino acid sequence of one of the five basic antigens; L is an optional linker amino acid sequence; A is an optional N-terminal amino acid sequence; B is an optional C-terminal amino acid sequence; and n is 2, 3, 4 or 5.

If a -X- moiety has a leader peptide sequence in its wild-type form, this may be included or omitted 10 in the hybrid protein. In some embodiments, the leader peptides will be deleted except for that of the -X- moiety located at the N-terminus of the hybrid protein *i.e.* the leader peptide of X_1 will be retained, but the leader peptides of $X_2 \dots X_n$ will be omitted. This is equivalent to deleting all leader peptides and using the leader peptide of X_1 as moiety -A-.

For each n instances of [-X-L-], linker amino acid sequence -L- may be present or absent. For 15 instance, when n=2 the hybrid may be $\text{NH}_2\text{-X}_1\text{-L}_1\text{-X}_2\text{-L}_2\text{-COOH}$, $\text{NH}_2\text{-X}_1\text{-X}_2\text{-COOH}$, $\text{NH}_2\text{-X}_1\text{-L}_1\text{-X}_2\text{-COOH}$, $\text{NH}_2\text{-X}_1\text{-X}_2\text{-L}_2\text{-COOH}$, *etc.* Linker amino acid sequence(s) -L- will typically be short (*e.g.* 20 or fewer amino acids *i.e.* 19, 18, 17, 16, 15, 14, 13, 12, 11, 10, 9, 8, 7, 6, 5, 4, 3, 2, 1). Examples 20 comprise short peptide sequences which facilitate cloning, poly-glycine linkers (*i.e.* comprising Gly_n where n = 2, 3, 4, 5, 6, 7, 8, 9, 10 or more), and histidine tags (*i.e.* His_n where n = 3, 4, 5, 6, 7, 8, 9, 10 or more). Other suitable linker amino acid sequences will be apparent to those skilled in the art. A 25 useful linker is GSGGGG (SEQ ID 9), with the Gly-Ser dipeptide being formed from a *Bam*HI restriction site, thus aiding cloning and manipulation, and the $(\text{Gly})_4$ tetrapeptide being a typical poly-glycine linker. If X_{n+1} is a ΔG protein and L_n is a glycine linker, this may be equivalent to X_{n+1} not being a ΔG protein and L_n being absent.

-A- is an optional N-terminal amino acid sequence. This will typically be short (*e.g.* 40 or fewer 25 amino acids *i.e.* 39, 38, 37, 36, 35, 34, 33, 32, 31, 30, 29, 28, 27, 26, 25, 24, 23, 22, 21, 20, 19, 18, 17, 16, 15, 14, 13, 12, 11, 10, 9, 8, 7, 6, 5, 4, 3, 2, 1). Examples include leader sequences to direct protein trafficking, or short peptide sequences which facilitate cloning or purification (*e.g.* histidine tags *i.e.* His_n where n = 3, 4, 5, 6, 7, 8, 9, 10 or more). Other suitable N-terminal amino acid 30 sequences will be apparent to those skilled in the art. If X_1 lacks its own N-terminus methionine, -A- is preferably an oligopeptide (*e.g.* with 1, 2, 3, 4, 5, 6, 7 or 8 amino acids) which provides a N-terminus methionine.

-B- is an optional C-terminal amino acid sequence. This will typically be short (*e.g.* 40 or fewer 35 amino acids *i.e.* 39, 38, 37, 36, 35, 34, 33, 32, 31, 30, 29, 28, 27, 26, 25, 24, 23, 22, 21, 20, 19, 18, 17, 16, 15, 14, 13, 12, 11, 10, 9, 8, 7, 6, 5, 4, 3, 2, 1). Examples include sequences to direct protein trafficking, short peptide sequences which facilitate cloning or purification (*e.g.* comprising histidine

tags *i.e.* His_n where $n = 3, 4, 5, 6, 7, 8, 9, 10$ or more), or sequences which enhance protein stability. Other suitable C-terminal amino acid sequences will be apparent to those skilled in the art.

Most preferably, n is 2. Two preferred proteins of this type are: X₁ is a 936 and X₂ is a 741; X₁ is a 287 and X₂ is a 953.

5 Two particularly preferred hybrid proteins of the invention are as follows:

n	A	X ₁	L ₁	X ₂	L ₂	B	[SEQ ID]
2	MA	ΔG287	GSGGGG	953 ^(NL)	—	—	7
2	M	936 ^(NL)	GSGGGG	ΔG741	—	—	8

These two proteins may be used in combination with NadA (particularly with SEQ ID 2).

936-ΔG741 hybrid can conveniently be prepared in highly pure form from expression in *E.coli* by a process comprising the steps of: homogenisation; centrifugation; cationic column chromatography; anionic column chromatography; hydrophobic column chromatography; diafiltration against a buffer;

10 and filter sterilisation. Further details of the process are given in the examples.

Sequences

The invention provides a polypeptide having an amino acid sequence selected from the group consisting of SEQ IDs 1 to 8. It also provides polypeptides having an amino acid sequence with sequence identity to an amino acid sequence selected from the group consisting of SEQ IDs 1 to 8.

15 As described above, the degree of 'sequence identity' is preferably greater than 50% (e.g. 60%, 70%, 80%, 90%, 95%, 99% or more).

The invention also provides nucleic acid encoding such polypeptides. Furthermore, the invention provides nucleic acid which can hybridise to this nucleic acid, preferably under "high stringency" conditions (e.g. 65°C in a 0.1xSSC, 0.5% SDS solution).

20 Polypeptides of the invention can be prepared by various means (e.g. recombinant expression, purification from cell culture, chemical synthesis (at least in part), *etc.*) and in various forms (e.g. native, fusions, non-glycosylated, lipidated, *etc.*). They are preferably prepared in substantially pure form (*i.e.* substantially free from other *N.meningitidis* or host cell proteins).

25 Nucleic acid according to the invention can be prepared in many ways (e.g. by chemical synthesis (at least in part), from genomic or cDNA libraries, from the organism itself, *etc.*) and can take various forms (e.g. single stranded, double stranded, vectors, probes, *etc.*). They are preferably prepared in substantially pure form (*i.e.* substantially free from other *N.meningitidis* or host cell nucleic acids).

The term "nucleic acid" includes DNA and RNA, and also their analogues, such as those containing modified backbones (e.g. phosphorothioates, *etc.*), and also peptide nucleic acids (PNA) *etc.* The

invention includes nucleic acid comprising sequences complementary to those described above (e.g. for antisense or probing purposes).

The invention also provides a process for producing a polypeptide of the invention, comprising the step of culturing a host cell transformed with nucleic acid of the invention under conditions which 5 induce polypeptide expression.

The invention provides a process for producing a polypeptide of the invention, comprising the step of synthesising at least part of the polypeptide by chemical means.

The invention provides a process for producing nucleic acid of the invention, comprising the step of amplifying nucleic acid using a primer-based amplification method (e.g. PCR).

10 The invention provides a process for producing nucleic acid of the invention, comprising the step of synthesising at least part of the nucleic acid by chemical means.

Strains

Preferred proteins of the invention comprise an amino acid sequence found in *N.meningitidis* serogroup B. Within serogroup B, preferred strains are 2996, MC58, 95N477, and 394/98. Strain 15 394/98 is sometimes referred to herein as 'NZ', as it is a New Zealand strain.

Protein 287 is preferably from strain 2996 or, more preferably, from strain 394/98.

Protein 741 is preferably from serogroup B strains MC58, 2996, 394/98, or 95N477, or from serogroup C strain 90/18311. Strain MC58 is more preferred.

Proteins 936, 953 and NadA are preferably from strain 2996.

20 Strains may be indicated as a subscript e.g. 741_{MC58} is protein 741 from strain MC58. Unless otherwise stated, proteins mentioned herein (e.g. with no subscript) are from *N.meningitidis* strain 2996, which can be taken as a 'reference' strain. It will be appreciated, however, that the invention is not in general limited by strain. As mentioned above, general references to a protein (e.g. '287', '919' etc.) may be taken to include that protein from any strain. This will typically have sequence 25 identity to 2996 of 90% or more (e.g. 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% or more).

Where hybrid proteins are used, the individual antigens within the hybrid (i.e. individual -X-moieties) may be from one or more strains. Where n=2, for instance, X₂ may be from the same strain as X₁ or from a different strain. Where n=3, the strains might be (i) X₁=X₂=X₃ (ii) X₁=X₂≠X₃ (iii) X₁≠X₂=X₃ (iv) X₁≠X₂≠X₃ or (v) X₁=X₃≠X₂, etc.

Hypervirulent lineages and bactericidal antibody responses

In general, compositions of the invention are able to induce serum bactericidal antibody responses after being administered to a subject. These responses are conveniently measured in mice and are a standard indicator of vaccine efficacy [e.g. see end-note 14 of reference 13].

5 Rather than offering narrow protection, compositions of the invention can induce bactericidal antibody responses against more than one hypervirulent lineage of serogroup B. In particular, they can induce bactericidal responses against two or three of the following three hypervirulent lineages: (i) cluster A4; (ii) ET5 complex; and (iii) lineage 3. They may additionally induce bactericidal antibody responses against one or more of hypervirulent lineages subgroup I, subgroup III, subgroup 10 IV-1 or ET-37 complex, and against other lineages e.g. hyperinvasive lineages.

This does not necessarily mean that the composition can induce bactericidal antibodies against each and every strain of serogroup B meningococcus within these hypervirulent lineages e.g. rather, for any given group of four or more strains of serogroup B meningococcus within a particular hypervirulent lineage, the antibodies induced by the composition are bactericidal against at least 50% 15 (e.g. 60%, 70%, 80%, 90% or more) of the group. Preferred groups of strains will include strains isolated in at least four of the following countries: GB, AU, CA, NO, IT, US, NZ, NL, BR, and CU. The serum preferably has a bactericidal titre of at least 1024 (e.g. 2^{10} , 2^{11} , 2^{12} , 2^{13} , 2^{14} , 2^{15} , 2^{16} , 2^{17} , 2^{18} or higher, preferably at least 2^{14}) i.e. the serum is able to kill at least 50% of test bacteria of a particular strain when diluted 1/1024, as described in reference 13.

20 Preferred compositions can induce bactericidal responses against the following strains of serogroup B meningococcus: (i) from cluster A4, strain 961-5945 (B:2b:P1.21,16) and/or strain G2136 (B:--); (ii) from ET-5 complex, strain MC58 (B:15:P1.7,16b) and/or strain 44/76 (B:15:P1.7,16); (iii) from lineage 3, strain 394/98 (B:4:P1.4) and/or strain BZ198 (B:NT:--). More preferred compositions can induce bactericidal responses against strains 961-5945, 44/76 and 394/98.

25 Strains 961-5945 and G2136 are both *Neisseria* MLST reference strains [ids 638 & 1002 in ref. 20]. Strain MC58 is widely available (e.g. ATCC BAA-335) and was the strain sequenced in reference 6. Strain 44/76 has been widely used and characterised (e.g. ref. 21) and is one of the *Neisseria* MLST reference strains [id 237 in ref. 20; row 32 of Table 2 in ref. 1]. Strain 394/98 was originally isolated in New Zealand in 1998, and there have been several published studies using this strain (e.g. refs. 22 30 & 23). Strain BZ198 is another MLST reference strain [id 409 in ref. 20; row 41 of Table 2 in ref. 1].

The composition may additionally induce a bactericidal response against serogroup W135 strain LNP17592 (W135:2a:P1.5,2), from ET-37 complex. This is a Haji strain isolated in France in 2000.

Heterologous host

Whilst expression of the proteins of the invention may take place in *Neisseria*, the present invention preferably utilises a heterologous host. The heterologous host may be prokaryotic (e.g. a bacterium) or eukaryotic. It is preferably *E.coli*, but other suitable hosts include *Bacillus subtilis*, *Vibrio cholerae*, *Salmonella typhi*, *Salmonella typhimurium*, *Neisseria lactamica*, *Neisseria cinerea*, *Mycobacteria* (e.g. *M.tuberculosis*), yeast, etc.

Thus the invention provides a composition which, after administration to a subject, is able to induce an antibody response in that subject, wherein the antibody response is bactericidal against two or more (e.g. 2 or 3) of hypervirulent lineages A4, ET-5 and lineage 3 of *N.meningitidis* serogroup B, and wherein the immunogens in the composition which give rise to the antibody response are obtained by recombinant expression in a non-Neisserial host. Thus the immunogens in the compositions of the invention are preferably recombinant immunogens. Compositions which do not include OMV preparations may thus be preferred.

Immunogenic compositions and medicaments

Compositions of the invention are preferably immunogenic compositions, and are more preferably vaccine compositions. The pH of the composition is preferably between 6 and 8, preferably about 7. pH may be maintained by the use of a buffer. The composition may be sterile and/or pyrogen-free.

Vaccines according to the invention may either be prophylactic (i.e. to prevent infection) or therapeutic (i.e. to treat infection), but will typically be prophylactic.

The invention also provides a composition of the invention for use as a medicament. The medicament is preferably able to raise an immune response in a mammal (i.e. it is an immunogenic composition) and is more preferably a vaccine.

The invention also provides the use of a composition of the invention in the manufacture of a medicament for raising an immune response in a mammal. It also provides the use of a 'NadA' protein, a '741' protein, a '936' protein, a '953' protein, and a '287' protein in the manufacture of a medicament for raising an immune response in a mammal. The medicament is preferably a vaccine.

The invention also provides a method for raising an immune response in a mammal comprising the step of administering an effective amount of a composition of the invention. The immune response is preferably protective and preferably involves antibodies. The method may raise a booster response.

The mammal is preferably a human. Where the vaccine is for prophylactic use, the human is preferably a child (e.g. a toddler or infant); where the vaccine is for therapeutic use, the human is preferably an adult. A vaccine intended for children may also be administered to adults e.g. to assess safety, dosage, immunogenicity, etc.

These uses and methods are preferably for the prevention and/or treatment of a disease caused by a *Neisseria* (e.g. meningitis, septicaemia, bacteremia, gonorrhoea etc.). The prevention and/or treatment of bacterial or meningococcal meningitis is preferred.

One way of checking efficacy of therapeutic treatment involves monitoring Neisserial infection after 5 administration of the composition of the invention. One way of checking efficacy of prophylactic treatment involves monitoring immune responses against the five basic antigens after administration of the composition.

Compositions of the invention will generally be administered directly to a patient. Direct delivery may be accomplished by parenteral injection (e.g. subcutaneously, intraperitoneally, intravenously, 10 intramuscularly, or to the interstitial space of a tissue), or by rectal, oral, vaginal, topical, transdermal, intranasal, ocular, aural, pulmonary or other mucosal administration.

The invention may be used to elicit systemic and/or mucosal immunity.

Dosage treatment can be a single dose schedule or a multiple dose schedule. Multiple doses may be used in a primary immunisation schedule and/or in a booster immunisation schedule.

15 Neisserial infections affect various areas of the body and so the compositions of the invention may be prepared in various forms. For example, the compositions may be prepared as injectables, either as liquid solutions or suspensions. Solid forms suitable for solution in, or suspension in, liquid vehicles prior to injection can also be prepared (e.g. a lyophilised composition). The composition may be prepared for topical administration e.g. as an ointment, cream or powder. The composition may be prepared for oral administration e.g. as a tablet or capsule, or as a syrup (optionally flavoured). The composition may be prepared for pulmonary administration e.g. as an inhaler, using a fine powder or a spray. The composition may be prepared as a suppository or pessary. The composition may be prepared for nasal, aural or ocular administration e.g. as drops.

20

25 Immunogenic compositions used as vaccines comprise an immunologically effective amount of antigen(s), as well as any other components, as needed. By 'immunologically effective amount', it is meant that the administration of that amount to an individual, either in a single dose or as part of a series, is effective for treatment or prevention. This amount varies depending upon the health and physical condition of the individual to be treated, age, the taxonomic group of individual to be treated (e.g. non-human primate, primate, etc.), the capacity of the individual's immune system to synthesise 30 antibodies, the degree of protection desired, the formulation of the vaccine, the treating doctor's assessment of the medical situation, and other relevant factors. It is expected that the amount will fall in a relatively broad range that can be determined through routine trials.

Further components of the composition

The composition of the invention will typically, in addition to the components mentioned above, 35 comprise one or more 'pharmaceutically acceptable carriers', which include any carrier that does not

itself induce the production of antibodies harmful to the individual receiving the composition. Suitable carriers are typically large, slowly metabolised macromolecules such as proteins, polysaccharides, polylactic acids, polyglycolic acids, polymeric amino acids, amino acid copolymers, and lipid aggregates (such as oil droplets or liposomes). Such carriers are well known to those of ordinary skill in the art. The vaccines may also contain diluents, such as water, saline, glycerol, etc. Additionally, auxiliary substances, such as wetting or emulsifying agents, pH buffering substances, and the like, may be present. A thorough discussion of pharmaceutically acceptable excipients is available in reference 24.

Vaccines of the invention may be administered in conjunction with other immunoregulatory agents. In particular, compositions will usually include an adjuvant. Preferred further adjuvants include, but are not limited to: (A) aluminium compounds (e.g. aluminium hydroxides, phosphates, aluminium, oxyhydroxide, orthophosphate, sulphate etc. [e.g. see chapters 8 & 9 of ref. 25]), or mixtures of different aluminium compounds, with the compounds taking any suitable form (e.g. gel, crystalline, amorphous etc.), and with adsorption being preferred; (B) MF59 (5% Squalene, 0.5% Tween 80, and 0.5% Span 85, formulated into submicron particles using a microfluidizer) [see Chapter 10 of 25; see also ref. 26]; (C) liposomes [see Chapters 13 and 14 of ref. 25]; (D) ISCOMs [see Chapter 23 of ref. 25], which may be devoid of additional detergent [27]; (E) SAF, containing 10% Squalane, 0.4% Tween 80, 5% pluronic-block polymer L121, and thr-MDP, either microfluidized into a submicron emulsion or vortexed to generate a larger particle size emulsion [see Chapter 12 of ref. 25]; (F) RibiTM adjuvant system (RAS), (Ribi Immunochem) containing 2% Squalene, 0.2% Tween 80, and one or more bacterial cell wall components from the group consisting of monophosphorylipid A (MPL), trehalose dimycolate (TDM), and cell wall skeleton (CWS), preferably MPL + CWS (DetoxTM); (G) saponin adjuvants, such as QuilA or QS21 [see Chapter 22 of ref. 25], also known as StimulonTM [28]; (H) chitosan [e.g. 29]; (I) complete Freund's adjuvant (CFA) and incomplete Freund's adjuvant (IFA); (J) cytokines, such as interleukins (e.g. IL-1, IL-2, IL-4, IL-5, IL-6, IL-7, IL-12, etc.), interferons (e.g. interferon- γ), macrophage colony stimulating factor, tumor necrosis factor, etc. [see Chapters 27 & 28 of ref. 25]; (K) monophosphoryl lipid A (MPL) or 3-O-deacylated MPL (3dMPL) [e.g. chapter 21 of ref. 25]; (L) combinations of 3dMPL with, for example, QS21 and/or oil-in-water emulsions [30]; (M) a polyoxyethylene ether or a polyoxyethylene ester [31]; (N) a polyoxyethylene sorbitan ester surfactant in combination with an octoxynol [32] or a polyoxyethylene alkyl ether or ester surfactant in combination with at least one additional non-ionic surfactant such as an octoxynol [33]; (O) a particle of metal salt [34]; (P) a saponin (e.g. QS21) + 3dMPL + IL-12 (optionally + a sterol) [36]; (Q) *E.coli* heat-labile enterotoxin ("LT"), or detoxified mutants thereof, such as the K63 or R72 mutants [e.g. Chapter 5 of ref. 37]; (R) cholera toxin ("CT"), or detoxified mutants thereof [e.g. Chapter 5 of ref. 37]; (S) double-stranded RNA; (T) microparticles (i.e. a particle of ~100nm to ~150 μ m in diameter, more preferably ~200nm to ~30 μ m in diameter, and most preferably ~500nm to ~10 μ m in diameter) formed from materials that are biodegradable and non-toxic (e.g. a poly(α -hydroxy acid), a polyhydroxybutyric acid, a polyorthoester, a polyanhydride, a polycaprolactone etc.), with

poly(lactide-co-glycolide) being preferred, optionally treated to have a negatively-charged surface (e.g. with SDS) or a positively-charged surface (e.g. with a cationic detergent, such as CTAB); (U) oligonucleotides comprising CpG motifs *i.e.* containing at least one CG dinucleotide, with 5-methylcytosine optionally being used in place of cytosine; (V) monophosphoryl lipid A mimics, such as aminoalkyl glucosaminide phosphate derivatives *e.g.* RC-529 [38]; (W) polyphosphazene (PCPP); (X) a bioadhesive [39] such as esterified hyaluronic acid microspheres [40] or a mucoadhesive selected from the group consisting of cross-linked derivatives of poly(acrylic acid), polyvinyl alcohol, polyvinyl pyrrolidone, polysaccharides and carboxymethylcellulose; or (Y) other substances that act as immunostimulating agents to enhance the effectiveness of the composition [e.g. see Chapter 7 of ref. 25]. Aluminium salts and MF59 are preferred adjuvants for parenteral immunisation. Mutant toxins are preferred mucosal adjuvants.

The use of an aluminium hydroxide adjuvant is particularly preferred.

Muramyl peptides include N-acetyl-muramyl-L-threonyl-D-isoglutamine (thr-MDP), N-acetyl-normuramyl-L-alanyl-D-isoglutamine (nor-MDP), N-acetylmuramyl-L-alanyl-D-isoglutaminyl-L-alanine-2-(1'-2'-dipalmitoyl-sn-glycero-3-hydroxyphosphoryloxy)-ethylamine MTP-PE), *etc.*

Further antigens

The composition contains five basic meningococcal protein antigens. It may also include further antigens, although it may contain no meningococcal protein antigens other than the five basic antigens. Further antigens for inclusion may be, for example:

- 20 - an outer-membrane vesicle (OMV) preparation from *N.meningitidis* serogroup B, such as those disclosed in refs. 4, 41, 42, 43 *etc.*
- a saccharide antigen from *N.meningitidis* serogroup A, C, W135 and/or Y, such as the oligosaccharide disclosed in ref. 44 from serogroup C [see also ref. 45] or the oligosaccharides of ref. 46.
- 25 - antigens from *Helicobacter pylori* such as CagA [47 to 50], VacA [51, 52], NAP [53, 54, 55], HopX [e.g. 56], HopY [e.g. 56] and/or urease.
- a saccharide antigen from *Streptococcus pneumoniae* [e.g. 57, 58, 59].
- an antigen from hepatitis A virus, such as inactivated virus [e.g. 60, 61].
- an antigen from hepatitis B virus, such as the surface and/or core antigens [e.g. 61, 62].
- 30 - an antigen from hepatitis C virus [e.g. 63].
- a diphtheria antigen, such as a diphtheria toxoid [e.g. chapter 3 of ref. 64] *e.g.* the CRM₁₉₇ mutant [e.g. 65].
- a tetanus antigen, such as a tetanus toxoid [e.g. chapter 4 of ref. 85].
- an antigen from *Bordetella pertussis*, such as pertussis holotoxin (PT) and filamentous 35 haemagglutinin (FHA) from *B.pertussis*, optionally also in combination with pertactin and/or agglutinogens 2 and 3 [e.g. refs. 66 & 67].
- a saccharide antigen from *Haemophilus influenzae* B [e.g. 45].

- polio antigen(s) [e.g. 68, 69] such as OPV or, preferably, IPV.
- an antigen from *N.gonorrhoeae* [e.g. 70, 71, 72, 73].
- an antigen from *Chlamydia pneumoniae* [e.g. refs. 74 to 80].
- an antigen from *Chlamydia trachomatis* [e.g. 81].
- 5 — an antigen from *Porphyromonas gingivalis* [e.g. 82].
- rabies antigen(s) [e.g. 83] such as lyophilised inactivated virus [e.g. 84, RabAvertTM].
- measles, mumps and/or rubella antigens [e.g. chapters 9, 10 & 11 of ref. 85].
- influenza antigen(s) [e.g. chapter 19 of ref. 85], such as the haemagglutinin and/or neuraminidase surface proteins.
- 10 — antigen(s) from a paramyxovirus such as respiratory syncytial virus (RSV [86, 87]) and/or parainfluenza virus (PIV3 [88]).
- an antigen from *Moraxella catarrhalis* [e.g. 89].
- an antigen from *Streptococcus pyogenes* (group A streptococcus) [e.g. 90, 91, 92].
- an antigen from *Staphylococcus aureus* [e.g. 93].
- 15 — an antigen from *Bacillus anthracis* [e.g. 94, 95, 96].
- an antigen from a virus in the flaviviridae family (genus flavivirus), such as from yellow fever virus, Japanese encephalitis virus, four serotypes of Dengue viruses, tick-borne encephalitis virus, West Nile virus.
- a pestivirus antigen, such as from classical porcine fever virus, bovine viral diarrhoea virus, and/or border disease virus.
- 20 — a parvovirus antigen e.g. from parvovirus B19.
- a prion protein (e.g. the CJD prion protein)
- an amyloid protein, such as a beta peptide [97]
- a cancer antigen, such as those listed in Table 1 of ref. 98 or in tables 3 & 4 of ref. 99.

25 The composition may comprise one or more of these further antigens.

Where a saccharide or carbohydrate antigen is used, it is preferably conjugated to a carrier protein in order to enhance immunogenicity [e.g. refs. 100 to 109]. Preferred carrier proteins are bacterial toxins or toxoids, such as diphtheria or tetanus toxoids. The CRM₁₉₇ diphtheria toxoid is particularly preferred [110]. Other carrier polypeptides include the *N.meningitidis* outer membrane protein [111], synthetic peptides [112, 113], heat shock proteins [114, 115], pertussis proteins [116, 117], protein D from *H.influenzae* [118], cytokines [119], lymphokines [119], hormones [119], growth factors [119], toxin A or B from *C.difficile* [120], iron-uptake proteins [121] etc. Where a mixture comprises capsular saccharides from both serogroups A and C, it is preferred that the ratio (w/w) of MenA saccharide:MenC saccharide is greater than 1 (e.g. 2:1, 3:1, 4:1, 5:1, 10:1 or higher). Saccharides from different serogroups of *N.meningitidis* may be conjugated to the same or different carrier proteins. Any suitable conjugation reaction can be used, with any suitable linker where necessary.

Toxic protein antigens may be detoxified where necessary (e.g. detoxification of pertussis toxin by chemical and/or genetic means [67]).

Where a diphtheria antigen is included in the composition it is preferred also to include tetanus antigen and pertussis antigens. Similarly, where a tetanus antigen is included it is preferred also to include diphtheria and pertussis antigens. Similarly, where a pertussis antigen is included it is preferred also to include diphtheria and tetanus antigens.

5 Antigens in the composition will typically be present at a concentration of at least 1 μ g/ml each. In general, the concentration of any given antigen will be sufficient to elicit an immune response against that antigen.

10 As an alternative to using protein antigens in the composition of the invention, nucleic acid encoding the antigen may be used [e.g. refs. 122 to 130]. Protein components of the compositions of the invention may thus be replaced by nucleic acid (preferably DNA e.g. in the form of a plasmid) that encodes the protein.

Definitions

15 The term "comprising" means "including" as well as "consisting" e.g. a composition "comprising" X may consist exclusively of X or may include something additional e.g. X + Y.

The term "about" in relation to a numerical value x means, for example, $x \pm 10\%$.

References to a percentage sequence identity between two amino acid sequences means that, when aligned, that percentage of amino acids are the same in comparing the two sequences. This alignment and the percent homology or sequence identity can be determined using software programs known in the art, for example those described in section 7.7.18 of reference 131. A preferred alignment is determined by the Smith-Waterman homology search algorithm using an affine gap search with a gap open penalty of 12 and a gap extension penalty of 2, BLOSUM matrix of 62. The Smith-Waterman homology search algorithm is taught in reference 132.

25 **MODES FOR CARRYING OUT THE INVENTION**

ΔG287-953 hybrid protein

DNA encoding protein 287 from meningococcal serogroup B strain 394/98 and protein 953 from meningococcal serogroup B strain 2996 were digested and ligated, together with a short linker sequence, to give a plasmid encoding amino acid sequence SEQ ID 7. The plasmid was transfected 30 into *E.coli* and bacteria were grown to express the protein.

After adequate growth, bacteria were harvested and the protein was purified. From culture, bacteria were centrifuged and the pellet was homogenized in the presence of 50 mM acetate buffer (pH 5) with a pellet:buffer volume ratio of 1: 8. Lysis was performed using a high pressure homogenizer

(AVESTIN, 4 cycles at 14000 psi). After lysis, urea was added at final concentration of 5M, followed by agitation for 1 hour at room temperature. The pH was reduced from 6 to 5 using 200 mM acetate buffer (pH 4)+ 5 M urea. The mixture was centrifuged at 16800g for 60 minutes at 2-8°C. The supernatant was collected and filtered by SARTOBRAN P (0.45-0.22µm SARTORIUS).

5 Protein in the filtered supernatant was stable for at least 30 days at -20°C and for at least 15 days at 2-8°C.

10 Protein was further purified on a cationic exchange column (SPFF, Amersham Biosciences) with elution using 350mM NaCl + 50 mM acetate + 5 M urea pH 5.00. The majority of impurities were present in the flow-thru. A pre-elution washing using a lower NaCl concentration (180 mM) advantageously eliminated two contaminating *E.coli* proteins .

The eluted material was adjusted to pH 8 (using 200 mM TRIS/HCl + 5 M urea pH 9) and further purified on a Q Sepharose HP column (Amersham) with elution using 150 mM NaCl + 20 mM TRIS/HCl pH 8.00 in 5 M urea. Again, a pre-elution washing with reduced salt (90 mM) was useful for eliminating impurities.

15 15 The filtered eluted material from Q HP column was diluted 1:2 using PBS pH 7.00 (150 mM NaCl + 10 mM potassium phosphate, pH 7.00) and then diafiltered against 10 volumes of PBS pH 7.00 by tangential ultrafiltration. At the end of diafiltration the material was concentrated 1.6 times to about 1.2 mg/ml total proteins. Using a 30,000 Da cut-off membrane (Regenerated Cellulose membrane 50cm², Millipore PLCTK 30) it was possible to dialyze the material with a yield of about 90% .

20 **936-ΔG741 hybrid protein**

25 DNA encoding protein 936 from meningococcal serogroup B strain 2996 and protein 741 from meningococcal serogroup B strain MC58 were digested and ligated, together with a short linker sequence, to give a plasmid encoding amino acid sequence SEQ ID 8. The plasmid was transfected into *E.coli* and bacteria were grown to express the protein. The recombinant protein was not secreted, but remained soluble within the bacteria.

After adequate growth, bacteria were centrifuged to give a humid paste and treated as follows:

- Homogenisation by high pressure system in presence of 20mM sodium phosphate pH 7.00.
- Centrifugation and clarification by orthogonal filtration.
- Cationic column chromatography (SP Sepharose Fast Flow), with elution by 150mM NaCl in 20mM sodium phosphate pH 7.00.
- Anionic column chromatography (Q Sepharose XL) with flow-through harvesting.
- Hydrophobic column chromatography (Phenyl Sepharose 6 Fast Flow High Sub) with elution by 20mM sodium phosphate, pH 7.00.
- Diafiltration against PBS pH 7.4 with a 10Kd cut-off.

— Final sterile filtration and storing at -20°C

Protein in the final material was stable for at least 3 months both at -20°C and at 2-8°C.

NadA^{(NL)(C)} protein

DNA encoding NadA protein from meningococcal serogroup B strain 2996 was digested to remove the sequence encoding its C-terminus, to give a plasmid encoding amino acid sequence SEQ ID 1. The plasmid was transfected into *E.coli* and bacteria were grown to express the protein. The recombinant protein was secreted into the culture medium, and the leader peptide was absent in the secreted protein (SEQ ID 2). The supernatant was treated as follows:

- Concentration 7X and diafiltration against buffer 20mM TRIS/HCl pH7.6 by cross flow UF (Cut off 30Kd).
- Anionic column chromatography (Q Sepharose XL), with elution by 400mM NaCl in 20mM TRIS/HCl pH 7.6.
- Hydrophobic column chromatography step (Phenyl Sepharose 6 Fast Flow High Sub), with elution by 50mM NaCl in TRIS/HCl pH 7.6.
- Hydroxylapatite ceramic column chromatography (HA Macro. Prep) with elution by 200mM sodium phosphate pH 7.4.
- Diafiltration (cut off 30Kd) against PBS pH 7.4
- Final sterile filtration and storing at -20°C

Protein in the final material was stable for at least 6 months both at -20°C and at 2-8°C.

NadA protein is susceptible to degradation, and truncated forms of NadA may be detected by western blot or by mass spectrometry (e.g. by MALDI-TOF) indicating up to 10kDa MW loss. Degradation products can be separated from native NadA by gel filtration (e.g. using column TSK 300SWXL, precolumn TSKSWXL, TOSOHAAS). Such filtration gives three peaks: (i) a first peak with retention time 12.637 min and apparent MW 885.036 Da; (ii) retention time 13.871 min and apparent MW 530.388 Da; (iii) retention time 13.871 min and apparent MW 530.388 Da. Light scattering analysis of the three peaks reveals real MW values of (i) 208500 Da, (ii) 98460 Da, (iii) 78760 Da. Thus the first peak contains NadA aggregates, and the third peak contains degradation products.

As the predicted molecular weight of NadA^{(NL)(C)} is 34.113 Da, peak (ii) contains a trimeric protein, which is the desired antigen.

30

Antigenic combinations

Mice were immunised with a composition comprising the three proteins and an aluminium hydroxide adjuvant. For comparison purposes, the three proteins were also tested singly. Ten mice were used per group. The mixture was able to induce high bactericidal titres against various strains:

	Meningococcal strain (Serogroup)							
	2996 (B)	MC58 (B)	NGH38	394/98 (B)	H44/76 (B)	F6124 (A)	BZ133 (C)	C11 (C)
(1)	32000	16000	130000	16000	32000	8000	16000	8000
(2)	256	131000	128	16000	32000	8000	16000	<4
(3)	32000	8000	—	—	—	8000	—	32000
Mix	32000	32000	65000	16000	260000	65000	>65000	8000

— indicates that this strain contains no NadA gene

Looking at individual mice, the triple mixture induced high and consistent bactericidal titres against the three serogroup B strains from which the individual antigens are derived:

#	1	2	3	4	5	6	7	8	9	10
2996	32768	16384	65536	32768	32768	65536	65536	32768	65536	8192
MC58	65536	32768	65536	65536	65536	8192	65536	32768	32768	65536
394/98	65536	4096	16384	4096	8192	4096	32768	16384	8192	16384

5

Combination and comparison with OMVs

10 In further experiments, the adjuvanted antigens (20 μ g of each antigen per dose) were administered in combination with 10 μ g OMVs prepared either from strain H44/76 (Norway) or strain 394/98 (New Zealand). Positive controls were the anti-capsular SEAM-3 mAb for serogroup B or CRM197-conjugated capsular saccharides for other strains. Results (bactericidal titres) are shown in Table 1. The mixture almost always gives better titres than simple OMVs and, furthermore, the addition of the mixture to OMVs almost always significantly enhances the efficacy of the OMVs. Moreover, in many cases the antigen mixture matches or exceeds the response seen with the positive control.

Hypervirulent lineage tests

15 The following antigens were tested against a variety of serogroup B strains from a variety of hypervirulent lineages:

- NadA^{(NL)(C)}
- Δ G287-953
- 936- Δ G741
- a mixture of (a), (b) and (c)
- OMVs prepared from strain H44/76 (Norway)
- OMVs prepared from strain 394/98 (New Zealand)
- A mixture of Δ G287 and (e)
- A mixture of (d) and (e)
- A mixture of (d) and (f)

20

25

SEAM-3 was used as a positive control.

Results were as follows, expressed as the percentage of strains in the indicated hypervirulent lineage where the serum bactericidal titre exceeded 1024:

	# strains	(a)	(b)	(c)	(d)	(e)	(f)	(g)	(h)	(i)	S-3
A4	4	50	50	0	100	25	25	25	100	100	+
ET-5	8	25	75	88	100	71	14	71	100	100	+
Lineage 3	13	0	75	15	93	8	85	8	92	93	+
ET-37	4	11	22	0	33	0	0	0	22	25	+

Against particular reference strains, bactericidal titres were as follows:

	Strain	(a)	(b)	(c)	(d)	(e)	(f)	(g)	(h)	(i)	S-3
A4	961-5945	128	2048	<8	2048	262144	8192	262144	262144	4096	8192
ET-5	44/76	<4	2048	32768	131072	524288	8192	524288	524288	524288	16384
Lineage 3	394/98	<4	1024	32	4096	<4	16384	256	16384	16384	16384
ET-37	LPN17592	2048	1024	256	4096	<8	<8	512	16384	65536	1024

5

Compositions (d), (h) and (i) therefore induce bactericidal antibody responses against a wide variety of strains of serogroup B meningococcus from within hypervirulent lineages A4, ET-5 and lineage 3. Titres using compositions (h) and (i) were generally higher than with (d), but the coverage of strains within hypervirulent lineages A4, ET-5 and lineage 3 were no better.

10 Coverage of untyped strains was also high with compositions (d), (h) and (i).

It will be understood that the invention has been described by way of example only and modifications may be made whilst remaining within the scope and spirit of the invention.

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CLAIMS

1. A composition comprising meningococcal antigens: (1) a 'NadA' protein; (2) a '741' protein; (3) a '936' protein; (4) a '953' protein; and (5) a '287' protein.
2. The composition of claim 1, wherein the NadA protein has 85% or more identity to SEQ ID 2.
- 5 3. The composition of claim 2, wherein the NadA protein comprises SEQ ID 2.
4. The composition of any preceding claim, wherein the 741 protein has 85% or more identity to SEQ ID 3.
5. The composition of claim 4, wherein the 741 protein comprises SEQ ID 3.
6. The composition of any preceding claim, wherein the 936 protein has 85% or more identity to SEQ ID 4.
- 10 7. The composition of claim 6, wherein the 936 protein comprises SEQ ID 4.
8. The composition of any preceding claim, wherein the 953 protein has 85% or more identity to SEQ ID 5.
9. The composition of claim 8, wherein the 953 protein comprises SEQ ID 5.
- 15 10. The composition of any preceding claim, wherein the 287 protein has 85% or more identity to SEQ ID 6.
11. The composition of claim 10, wherein the 287 protein comprises SEQ ID 6.
12. The composition of any preceding claim, wherein at least two of the antigens (1) to (5) are expressed as a single polypeptide chain.
- 20 13. The composition of claim 12, wherein the composition comprises a polypeptide which comprises a pair of antigens within a single polypeptide chain selected from the group consisting of: NadA & 741; NadA & 936; NadA & 953; NadA & 287; 741 & 936; 741 & 953; 741 & 287; 936 & 953; 936 & 287; 953 & 287.
- 25 14. The composition of claim 12 or claim 13, wherein the composition comprises a polypeptide of formula $\text{NH}_2\text{-A-}[-\text{X-L-}]_n\text{-B-COOH}$, wherein: X is an amino acid sequence of one of the five antigens (1) to (5); L is an optional linker amino acid sequence; A is an optional N-terminal amino acid sequence; B is an optional C-terminal amino acid sequence; and n is 2, 3, 4 or 5.
15. The composition of claim 14, wherein n is 2, X_1 is a 936 protein and X_2 is a 741 protein.
16. The composition of claim 14, wherein n is 2, X_1 is a 287 protein and X_2 is a 953 protein.

17. The composition of any preceding claim, comprising a protein comprising SEQ ID 7.
18. The composition of any preceding claim, comprising a protein comprising SEQ ID 8.
19. A composition which, after administration to a subject, is able to induce an antibody response in that subject, wherein the antibody response is bactericidal against two or more of hypervirulent lineages A4, ET-5 and lineage 3 of *N.meningitidis* serogroup B.
5
20. The composition of claim 19, wherein the components which give rise to the bactericidal antibody response are obtained by recombinant expression.
21. The composition of any preceding claim, for use as a medicament.
22. The use of a composition of any preceding claim in the manufacture of a medicament for the prevention and/or treatment of a disease caused by a *Neisseria*.
10
23. A method for raising an antibody response in a mammal, comprising the step of administering an effective amount of a composition according to any one of claims 1 to 20.
24. A polypeptide having an amino acid sequence selected from the group consisting of SEQ IDs 1 to 8.
- 15 25. A process for purifying soluble NadA from a culture medium, comprising the steps of: concentration and diafiltration against a buffer by ultrafiltration; anionic column chromatography; hydrophobic column chromatography; hydroxylapatite ceramic column chromatography; diafiltration against a buffer; and filter sterilisation.
26. A process for purifying a 936-ΔG741 hybrid protein from a bacterium, comprising the steps of: homogenisation; centrifugation; cationic column chromatography; anionic column chromatography; hydrophobic column chromatography; diafiltration against a buffer; and filter sterilisation.
20

SEQUENCE LISTING

SEQ ID 1 – NadA from strain 2996, with C-terminus deletion

5 MKHFPSKVLTTAILATFCGALAAATNDDVKAATVAIAAAYNNQEINGFKAGETIYDIDEDEGTITKKDATAADVEADDFKGLKKVVTNLTKTVNENKQNVDAVKAAESEIEKLTTLADTDAALADTDAALDTNALKLGENITTFAEETKTNIV
KIDEKLEAVADTVDKHAEAFNDIADSLDETNKADEAVKTANEAKQTAETKQNVDAVKAAETAAGKAEAAAGTANTAAD
KAEAVAAKVTDIKADIATNKDNIACKKANSADVYTREESDSKFVRIDGLNATTEKLDTRLASAEKSIADHDTRLNGLDKTVSDLRKETRQGLAEQAALSGLFQPYNVG

SEQ ID 2 – NadA from strain 2996, with C-terminus deletion and leader peptide processed

10 ATNDDDVKAATVAIAAAYNNQEINGFKAGETIYDIDEDEGTITKKDATAADVEADDFKGLKKVVTNLTKTVNENKQNV
DAVKAAESEIEKLTTLADTDAALADTDAALDTNALKLGENITTFAEETKTNIVKIDEKLEAVADTVDKHAEAFNDI
ADSLDETNKADEAVKTANEAKQTAETKQNVDAVKAAETAAGKAEAAAGTANTAADKAEAVAAKVTDIKADIATNKDNI
ACKKANSADVYTREESDSKFVRIDGLNATTEKLDTRLASAEKSIADHDTRLNGLDKTVSDLRKETRQGLAEQAALSGLFQPY
NVG

SEQ ID 3 – AG741 from MC58 strain

15 VAADIGAGLADALTAPLDHKDKGLQSLTLQSVRKNEKLKLAAGQGAEKTYGNGDSLNTGKLKNDKVSREFDIFRQIEVDGQL
ITLESGEFQVYKQSHSALTAFQTEQIQDSEHSGKMVKRQFRIGDIAGEHTSFDFLPEGGRATYRGTAFGSDDAGGKLTYT
IDFAAKQGNGKIEHLKSPELNVDLAAADIKPDGKRHAVISGSVLYNQAEKGSYSLGI FGGKAQEVAEVKTVNGIRHIG
LAAKQ

SEQ ID 4 – 936 from MC58 strain with leader peptide processed

20 VSAVIGSAAVGAKSAVDRRTTGAQTDDNVMALRIETTARSYLRQNNQTKGYTPQISVVGYNRHLLLQVATEGEKQFVGQ
IARSEQAAEVVNYITVASLPRTAGDIAGDTWNTSKVRATLLGISPATQARVKIVTYGNVTYVMGILTPEEQAQITQKVST
TVGVQKVITLYQNYVQR

SEQ ID 5 – 953 from MC58 strain with leader peptide processed

25 ATYKVDEYHANARFAIDHFNTSTNVGGFYGLTGSVEFDQAKRDGKIDITIPIANLQSGSQHFTDHLKSADI FDAAQYPDIR
FVSTKFNFNGKKLVSVDGNLTMHGKTAPVKLAEKFNCYQSPMEKTEVCGGDFSTTIDRTKWGMDYLVNVGMTKSVRIDIQ
IEAAKQ

SEQ ID 6 – AG287 from MC58 strain

30 SPDVKSAADTLSPKAAPVSEKETEAKEDAPQAGSQGQGAPSQGSQDMAAVSEENTNGGAVTADNPNEDEVAQNDMPQN
AAGTDSSTPNHTPDPNMLAGNMENQATDAGESSQPANQPDMANAADGMQGDDPSAGGQNAGNTAAQGANQAGNNQAAGSSD
PIPASNPAPANGGSNFGRVDLANGVLIDGPSQNITLTHCKGDSCSGNNFLDEEVQLKSEFEKLSADKISNYKKDGKNDKF
VGLVADSVQMKGINQYIIFYKPKPTSFAFRRSARSRRSLPAEMPLIPVNQADTLIVDGEAVSLTGHSGNI FAPEGNYRYL
TYGAEKLPGGSYALRVQGEPAKGEMLAGAAVYNGEVLFHTENGRPYPTRGRFAAKVDFGSKSVDGIIDSGDDLHMGQTQKF
KAAIDNGNGFKGTWTENGSDVSGKFYGPAGEEVAGKSYRPTDAEKGGFGVFAGKKEQD

SEQ ID 7 – 287-953 hybrid

35 MASPDVKSAADTLSPKAAPVSEKETEAKEDAPQAGSQGQGAPSQGSQDMAAVSEENTNGGAAATDKPKNEDEGAQNDMPPM
QNAADTDSLTPNHTPASNMPAGNMENQAPDAGESQPANQPDMANAADGMQGDDPSAGGQNAGNTAAQGTNQAENNQTAGS
QNPASSTNPSATNSGGDFGRNVGNSSVVIDGPSQNITLTHCKGDSCSGNNFLDEEVQLKSEFEKLSADKISNYKKDGKND
GKNDKFVGLVADSVQMKGINQYIIFYKPKPTSFAFRRSARSRRSLPAEMPLIPVNQADTLIVDGEAVSLTGHSGNI FAPE
GNYRYLTYGAEKLPGGSYALRVQGEPAKGEMLAGTAVYNGEVLFHTENGRPSPSRGRFAAKVDFGSKSVDGIIDSGDGLH
40 MGTQKFKAIDGNGFKGTWTENGSDVSGKFYGPAGEEVAGKSYRPTDAEKGGFGVFAGKKEQDGSGGGGATYKVDEYHA
NARFAIDHFNTSTNVGGFYGLTGSVEFDQAKRDGKIDITIPVANLQSGSQHFTDHLKSADI FDAAQYPDIRFVSTKFNFNG
KKLVSVDGNLTMHGKTAPVKLAEKFNCYQSPMAKTEVCGGDFSTTIDRTKWGVDYLVNVGMTKSVRIDIQIEAAKQ*

SEQ ID 8 – 936-741 hybrid

MVSAIGSAAVGAKSAVDRRTGAGTDDNVMALRIETTARSYLRQNNQTKGYTPQISVVGYNRHLLLLGQVATEGEKQFVG
QIARSEQAAEGVNYITVASLPRTAGDIAGDTWNTSKVRATLLGISPATQARVKIVTYGNVTYVMGILTPEEQAQITQKVS
TTVGVQKVITLYQNYVQRGSGGGVAAAGAGLADALTAPLDHKDKGLQSLTLDQSVRKNEKLKLAQGAEKTYGNGDSLNTGKLKNDKVSRFDFIRQ
5 TGKLKNDKVSRFDFIRQIEVDGQLITLESGEFQVYKQSHSALTAFQTEQIQLDSEHSGKMVAKRQFRIGDIAGEHTSFDKLP
EGGRATYRGTAFGSDDAGGKLTYTIDFAAKQGNGKIEHLKSPELNVDLAAADIKPDGKRHAVISGSVLYNQAEKGSYSLGI
FGGKAQEVAEVKTVNGIRHIGLAAKQ*

SEQ ID 9 – linker

GSGGGG

SEQ ID 10

CSSGGGGVAADIGAGLADALTAPLDHKDKGLQSLTLDQSVRKNEKLKLAQGAEKTYGNGDSLNTGKLKNDKVSRFDFIRQ
IEVDGQLITLESGEFQVYKQSHSALTAFQTEQIQLDSEHSGKMVAKRQFRIGDIAGEHTSFDKLP
GGKLTYTIDFAAKQGNGKIEHLKSPELNVDLAAADIKPDGKRHAVISGSVLYNQAEKGSYSLGI
NGIRHIGLAAKQ

SEQ ID 11

CSSGGGGVAADIGAGLADALTAPLDHKDKSLQSLTLDQSVRKNEKLKLAQGAEKTYGNGDSLNTGKLKNDKVSRFDFIRQ
IEVDGQLITLESGEFQIYKQDHSAVVALQIEKINNPDKIDS LINQRSFLVSGLGGEHTAFNQLPDGKA
6 GKLTYTIDFAAKQGHGKIEHLKTPEQNV
ELAAAEKSHAVILGDTRYGSEEKGYH
LA
LG
DRA
Q
E
IAG
KVHEIGIAGKQ

SEQ ID 12

CSSGGGGSGGGGVAADIGTGLADALTAPLDHKDKGLKSLTLEDSIPQNGTLTL
SAQGAEKTFKAGDKDNSLNTGKLKNDK
SRDFVQKIEVDGQTITLASGEFQIYKQNSAVVALQIEKINNPDKTDS LINQRSFLVSGLGGEHTAFNQLPGGKA
AFSSDDPNGR
LHYSIDFTKQGYGRIEHLKTLEQNV
ELAAAEKSHAVILGDTRYGSEEKGYH
LA
LG
DRA
Q
E
IAG
SATVKIGEKVHEIGIAGKQ

TABLE 1

	2996	NGH38	M4215	MC58	4476	CU385	N44/89	394/98	M01-240149	NM008	BZ198	961-5945	G2136	5/99	F6/124	BZ133	LPN11592	240539
Typing:	B:15:P1.5a	B:NT: P1.3	B:15:P1.7,16	B:15:P1.7,16	B:15:P1.7,16	B:4:P1.15	B:4:P1.15	B:4:P1.15	B:4:P1.7,4	B:4:P1.4	B:NT	B:2b:P1.21,16	B:	B:2b:P1.5,2	A	C:NT:	W135	P1.5
ET:	other	other	n.d.	ET5	ET5	ET5	lin.3	lin.3	lin.3	lin.3	A4	A4	A4	A4	sII	sI	o	
Positive control	32768	32768	32768	16384	16384	>16384	8192	16384	8192	32768	8192	16384	8192	32768	8192	1024	1024	4096
Antigen mixture	4096	4096	65536	32768	65536	>65536	>4096	8192	2048	>4096	4096	2048	2048	>4096	8192	16384	4096	>8192
Antigens + H4476 OMVs	16384	8192	>65536	32768	524288	>65536	>4096	16384	8192	>4096	>4096	>8192	2048	>4096	8192	32768	16384	>8192
Antigens + 394/98 OMVs	8192	8192	>65536	32768	>65536	>65536	>4096	65536	>8192	>4096	>4096	>8192	2048	>4096	65536	32768	16384	>8192
OMVs (Norway)	<4	1024	8192	2048	262144	256	<8	4096	<4	<8	<4	>8192	<8	<8	<8	1024	<4	<8
OMVs (NZ)	512	<4	128	2048	<4	<8	<8	32768	>8192	4096	1024	4096	<16	n.d.	<8	4096	1024	<8

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